

Apoe Targeted Mutation Mice

Inactivation of the Apoe gene provides a mouse model for human atherogenesis and diverse applications in studies of lipid transport and metabolism

Applications for the Apoe Targeted Mutation Mouse Model

The lack of a functional *Apoe* gene renders these mice unable to produce a key glycoprotein, apoE (apolipoprotein E), which is essential for the transport and metabolism of lipids. The mice are healthy when born, but have a markedly altered plasma lipid profile compared to normal mice, and rapidly develop atherosclerotic lesions. Numerous potential applications to basic research and drug development include:

- Cholesterol/triglyceride transport mechanisms
- Role of apoE in lipoprotein binding to endothelium and the incorporation of lipids into atherosclerotic plaques
- Cholesterol reverse transport
- Relative contributions of dietary constituents to atherogenesis
- Modeling of human apoE-deficiency
- Impact of lipid oxidation on cellular lipid uptake/metabolism
- Role of apoA-I and apoA-IV in lipoprotein formation and transport
- Apoe function in normal and diseased neural tissue and nerve repair
- Gene therapy to replace specific *Apoe* alleles in Apoe Targeted Mutation Mice
- Evaluation of pharmacologic manipulations of atherosclerotic plaque formation
- Development through crossbreeding of models for specific research programs in atherosclerosis and CNS disease

Features of the Apoe Targeted Mutation Mouse (model APOE)^{1,2}

- Homozygous for defective *Apoe* gene
- Complete absence of apoE glycoprotein

Lipoprotein Profiles of Normal and Apoe Targeted Mutation Mice

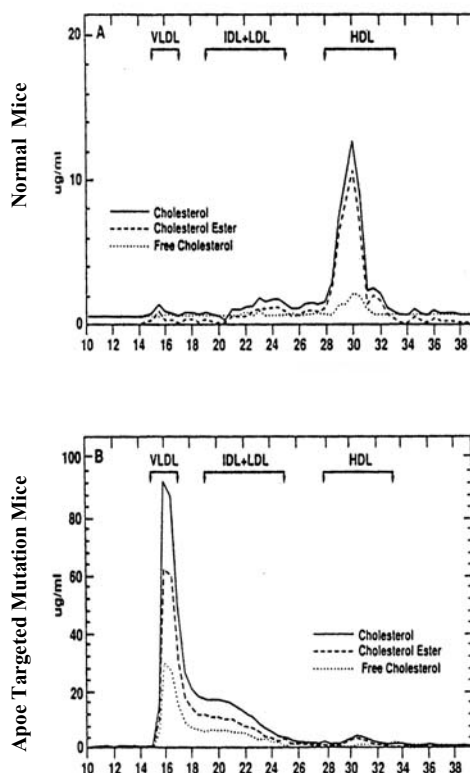


Figure 1. Reprinted with permission from the *Journal of Clinical Investigation*. Kashyap, et al.⁴

- Markedly elevated plasma cholesterol levels (5 times normal)
- Relatively normal plasma triglyceride levels
- Shift from the normal distribution of plasma lipids (cholesterol and triglycerides) from predominantly high density lipoprotein (HDL) complexes to predominately very low density lipoprotein (VLDL) and intermediate density lipoprotein (IDL) complexes²
- Early and severe spontaneous development of atherosclerotic lesions

- Normal reproductive performance in both male and female ApoE Targeted Mutation Mice²
- No reported difference in body weights or litter size compared to normal mice
- Enrichment of VLDL and IDL particles with apoA-I, apoA-IV, and apoB-48, similar to their distribution in humans who are apoE deficient, and similar to chylomicron remnants²

Scientific Profile of the ApoE Targeted Mutation Mouse Model

ApoE Targeted Mutation Mice are homozygous for a defective *ApoE* gene, and apoE protein is absent from blood of these animals.¹ Plasma lipoproteins in which apoE normally is an integral part (e.g., VLDL, IDL, HDL) are therefore devoid of this apolipoprotein.² As a result, ApoE Targeted Mutation Mice show dramatic alterations in lipid metabolism and transport, even on a normal mouse chow diet.

In normal mice, the protein apoE is made predominantly by the liver (as in humans) and becomes incorporated into all lipoprotein particles (except LDL), including those constructed by intestinal cells (chylomicrons) for transport of ingested lipids through the bloodstream, and those constructed by liver cells to distribute lipids via the circulation and fuel cellular metabolism during fasting (VLDLs).³ apoE is present in other lipoprotein particles that are metabolic products (IDLs, chylomicron remnants), or those involved in acquisition of cholesterol from tissues and reverse cholesterol transport (HDLs).

The most well-documented function of apoE is to facilitate binding of lipoproteins to cell surface receptors, thereby enhancing transfer of components of the particle's lipid moiety (e.g., cholesteryl esters, triacyl-glycerols) to or from cells. Specifically, apoE mediates binding to both LDL and LRP (LDL receptor-related protein) receptors, and possibly the VLDL receptor.⁴ Less well understood but intriguing are possible roles for apoE in nerve regeneration and amyloid processing in CNS neurons.

The absence of apoE has multiple effects on lipid metabolism and transport. In normal mice, the profile of cholesterol-containing plasma lipoproteins includes an abundance of high density lipoprotein (HDL) and only trace amounts of lower-density lipoproteins (VLDL, IDL, LDL).^{2,3} In contrast, ApoE Targeted Mutation Mice have the reverse distribution: 80% of serum cholesterol is sequestered in lower density lipoproteins, while HDL-cholesterol is half the normal level.² In addition, total cholesterol levels in plasma of ApoE Targeted Mutation mice are up to five times that of normal mice (e.g., 434 mg/dl versus 86 mg/dl, respectively).² Thus, both the quantity of cholesterol and its distribution among lipoprotein fractions are shifted in the mutant mice to a pattern known to be associated in humans with risk of atherosclerosis.

The alterations generated in the ApoE Targeted Mutation Mice confer a spontaneous susceptibility to early and extensive development of atheromas regardless of diet. Unlike normal mice, which do not develop atherosclerotic lesions (except for some strains on high fat/high cholesterol diets), ApoE Targeted Mutation Mice fed normal mouse chow develop foam cell accumulations on their aortic walls by 3 months of age, which progress to extensive atherosclerotic lesions and severe vessel occlusion by 8 months.²

Other than lipid alterations and propensity to develop atherosclerosis, ApoE Targeted Mutation Mice are of similar weight to normal mice and appear otherwise healthy.² This, combined with early atherosclerosis and underlying changes in plasma lipid profile that mimic the human atherogenic profile, make ApoE Targeted Mutation Mice an ideal model for human atherogenesis. They also can serve as a model for human apoE deficiency, a rare genetic disorder.

The apoE deficiency has been corrected experimentally in ApoE Targeted Mutation Mice, with a return to normal lipoprotein distribution and cholesterol levels (650 mg/dl in apoE deficient mice versus 100-150 mg/dl in apoE corrected mice).⁴ This was accomplished by gene replacement through infusion of an

adenovirus vector containing the human *ApoE* gene, and resulted in mice with markedly reduced incidence of atherosclerosis.

Origin of the Model

The ApoE Targeted Mutation Mouse Model was developed by Nobuyo Maeda and colleagues at the University of North Carolina in Chapel Hill.¹ Inactivation of the *ApoE* gene was accomplished by homologous recombination, by electroporation of cultured E14TG2a cells with a plasmid that contains a modification of the murine *ApoE* locus, in which a portion of the normal *ApoE* gene sequence had been replaced by neomycin-resistance and thymidine kinase marker genes.

Electroporated cells that had undergone successful recombination of wild-type and plasmid-derived *ApoE* gene sequences were identified by positive neomycin resistance, and hence, presence of the disrupted *ApoE* gene. Chimeric mice then were generated by injecting such cells into blastocysts of C57BL/6J mice.

Of the mice that developed from the blastocysts, a male chimera (JH126.1) was found to transmit the embryonic cell genome to 100% of his offspring in a backcross to a C57BL/6 female. The resulting pups inherited the *ApoE* gene in expected Mendelian proportion (50%), indicating integration of the mutation into the genome and no effect of the mutation on embryo viability.

The mice were transferred to Taconic in May 1998 after ten backcrosses (N10) to C57BL/6J from the (129 X C57BL/6) chimera and intracrossing to homozygosity. The line was then further backcrossed at Taconic to C57BL/6N (N11) and derived by embryo transfer. The colony is maintained at N11 through intercrossing of homozygous mice.

Ready for Your Experiments

Taconic's ApoE targeted mutation mice are produced in Isolator Barrier Unit (IBUTM) facilities. Mice are shipped in Taconic Transport Cages (TTCTM) and come with an up-to-date health report documenting their Murine

Pathogen Free (MPFTM) health status. Barrier housing conditions are recommended for maintenance of ApoE Targeted Mutation homozygous mice.

Related Mouse Models from Taconic

Taconic provides a number of mouse models relevant to lipoprotein metabolism and atherogenesis. Call, fax, or visit Taconic's website for information about these additional models:

- **ApoB100 Microinjected Mouse (model 001004)** – develops atherosclerosis on a high fat/high cholesterol diet, due to expression of high levels of human apolipoprotein B100 and a resultant elevated plasma cholesterol (total and LDL-associated).
- **APOE2 Targeted Replacement Mouse (model 001547)** – expressing the human apoE2 protein instead of murine apoE, with several abnormalities of lipid physiology, including elevated serum levels, altered lipoprotein profiles, and early development of atherosclerosis, all of which parallel features of human type III lipoproteinemia.
- **APOE3 Targeted Replacement Mouse (model 001548)** – expressing the human apoE3 protein instead of murine apoE, with normal serum cholesterol and triglyceride levels, but certain abnormalities of lipid physiology, including delayed clearance of lipoprotein particles (VLDL) and propensity to develop atherosclerosis on a high-fat diet.
- **APOE4 Targeted Replacement Mouse (model 001549)** – expressing the human apoE4 protein instead of murine apoE, with normal serum cholesterol and triglyceride levels but certain abnormalities of lipid physiology that are similar to those of ApoE3 Targeted Replacement Mice; impairment in clearance of lipoprotein particles (VLDL) and development of atherosclerosis on a high-fat diet are more pronounced.
- **CETP Microinjected Mouse (model 001003)** – expressing human cholesteryl ester transfer protein, a plasma enzyme normally absent in mice, which in humans mediates HDL-cholesterol transfer to VLDL

and LDL and enhances cholesterol uptake into cells; transgenic mice have dramatically reduced HDL-cholesterol.

- **CETP/ApoB100 Double Microinjected Mouse (model 001007)** – develops atherosclerosis rapidly on a high-fat/high-cholesterol diet or normal mouse chow after six months, due to expression of both human CETP and apoB100 and consequent alterations in plasmalipid profile (elevated LDL-cholesterol, reduced HDL-cholesterol)

References Cited

1. Piedrahita J.A., Zhang S.H., Hagaman J.R., Oliver P.M., Maeda N. (1992) **Generation of Mice Carrying a Mutant Apolipoprotein E Gene Inactivated by Gene Targeting in Embryonic Stem Cells.** *Proceedings of the National Academy of Science, USA*, Vol. 89, pp. 4471-4475.
2. Zhang S.H., Reddick R.L., Piedrahita J.A., Maeda N. (1992) **Spontaneous Hypercholesterolemia and Arterial Lesions in Mice Lacking Apolipoprotein E.** *Science*, Vol. 258, pp. 468-471
3. Plump A.S., Breslow J.L. (1995) **Apolipoprotein E and the Apolipoprotein E-Deficient Mouse.** *Annual Review of Nutrition*, Vol. 15, pp. 495-518.
4. Kashyap V.S., Santamarina-Fojo S., Brown D.R., Parrott C.L., Applebaum-Bowden D., Meyn S., Talley G., Paigen B., Maeda N., Brewer, Jr., H.B. (1995) **Apolipoprotein E Deficiency in Mice: Gene Replacement and Prevention of Atherosclerosis Using Adenovirus Vectors.** *Journal of Clinical Investigation*, Vol. 96, pp. 1612-1620.

© Copyright 2008, Taconic Farms, Inc. RG290495
TACONIC TRANSGENIC MODELS are produced and distributed under rights to patents and intellectual property licensed from various institutions. Transgenic Models are produced and distributed under United States Patent Nos. 4,873,191; 4,740,470; 5,464,764; 5,487,992; 5,627,059; 5,631,153; 5,789,215; and 6,204,061.

Every Taconic Transgenic Model™ carries a label license granting you a license under Taconic's in-licensed patent right(s) to use the model in your research. TTM™s are produced and distributed under rights to patents that Taconic has licensed from various institutions, including exclusive distribution rights to Positive Negative Selection and Isogenic DNA gene targeting technologies. Taconic is the only commercial breeder that can supply transgenic models with these licenses for use in your research.

Conditions of Use for Taconic Transgenic Models™

TACONIC TRANSGENIC MODELS™ (“MODELS”) are produced and distributed under rights to patents and intellectual property licensed from various institutions. Taconic grants to each purchaser a right under Taconic's rights in such licensed patents and intellectual property to use the purchased MODEL in consideration of purchasers' acknowledgement of and agreement to the Terms and Conditions of Sale and the following terms of use:

- Title to these MODELS and biological materials derived from them remains WITH TACONIC FARMS, INC.
- The MODELS will be used for research purposes only.

- The MODELS will not be bred except to obtain embryos or fetuses required for research purposes.
- The MODELS and biological materials derived from them will not be distributed to third parties or used for commercial purposes.

For more information or to place an order contact:

TACONIC
1 Discovery Drive, Suite 304
Rensselaer, NY 12144
Toll Free: 1-888-TACONIC
Phone: 518-537-6208
Fax: 518-537-7287
e-mail: custserv@taconic.com
Internet: <http://www.taconic.com>

in Europe: Taconic Europe
Bomholtvej 10 P.O. Box 39
DK 8680 Ry DENMARK
Phone: +45 70 23 04 05
Fax: +45 86 84 16 99
e-mail: TaconicEurope@taconic.com
Internet: <http://www.taconic.com>

in Japan: CLEA Japan, Inc.
Phone: 03-5704-7063
Fax: 03-3792-5298
e-mail: ad-import@clea-japan.com
Internet: <http://clea-japan.com>

Rev. 3/08

Please Note: e-mail transmission of this document may result in the loss of formatting or symbols, i.e., Greek letters or symbols for trademark, degrees, etc.

Taconic Transgenic Models Publication Reference List ApoE Targeted Mutation Mice

- Aihara K, Azuma H, Akaike M, Ikeda Y, Sata M, Takamori N, Yagi S, Iwase T, Sumitomo Y, Kawano H, Yamada T, Fukuda T, Matsumoto T, Sekine K, Sato T, Nakamichi Y, Yamamoto Y, Yoshimura K, Watanabe T, Nakamura T, Oomizu A, Tsukada M, Hayashi H, Sudo T, Kato S, Matsumoto T. (2007) **Strain-dependent embryonic lethality and exaggerated vascular remodeling in heparin cofactor II-deficient mice.** *Journal of Clinical Investigation*, 117(6):1514–26.
- Burgos JS, Ramirez C, Sastre I, Valdivieso F. (2006) **Effect of Apolipoprotein E on the Cerebral Load of Latent Herpes Simplex Virus Type 1 DNA.** *Journal of Virology*, 80(11):5383-7.
- Daugherty, A., Pure, E., Delfel-Butteiger, D., Chen, S., Lefterovich, J., Roselaar, S.E., Rader, D.J. (1997) **The Effects of Total Lymphocyte Deficiency on the Extent of Atherosclerosis in Apolipoprotein E-I- Mice,** *Journal of Clinical Investigation*, Vol. 100, No. 6, pp. 1575-1580.
- Guevara, N.V., Kim, H-S., Antonova, E.I., Chan, L. (1999) **The Absence of p53 Accelerates Atherosclerosis by Increasing Cell Proliferation in vivo,** *Nature Medicine*, Vol. 5, No. 3, pp. 335-339.
- Hodgin, J.B., Kregge, J.H., Reddick, R.L., Korach, K.S., Smithies, O., Maeda, N. (2001) **Estrogen Receptor Alpha is a Major Mediator of 17beta-estradiol's Atheroprotective Effects on Lesion Size in Apoe^{-/-} Mice,** *Journal of Clinical Investigation*, Vol. 107, No. 3, pp. 333-340.
- Ishibashi S, Herz J, Maeda N, Goldstein JL, Brown MS. (1994) **The two-receptor model of lipoprotein clearance: Tests of the hypothesis in "knockout" mice lacking the low density lipoprotein receptor, apolipoprotein E, or both proteins;** *Proc Natl Acad Sci*, 91:4431-5.
- Knowles JW, Reddick RL, Jennette JC, Shesely EG, Smithies O, Maeda N. (2000) **Enhanced atherosclerosis and kidney dysfunction in eNOS^{??} ApoE^{??} mice are ameliorated by enalapril treatment;** *J Clin Invest*, 105(4):451-8.
- Kashyap, V.S., Santamarina-Fojo, S., Brown, D.R., Parrott, C.L., Applebaum-Bowden, D., Meyn, S., Talley, G., Paigen, B., Maeda, N., Brewer, Jr., H.B. (1995) **Apolipoprotein E Deficiency in Mice: Gene Replacement and Prevention of Atherosclerosis Using Adenovirus Vectors,** *Journal of Clinical Investigation*, Vol. 96, pp. 1612-1620.
- Lippmann M, Ito K, Hwang JS, Maciejczyk P, Chen LC. (2006) **Cardiovascular Effects of Nickel in Ambient Air.** *Environmental Health Perspective*, 114(11):1662-9.
- Maezawa I, Maeda N, Montine TJ, Montine KS. (2006) **Apolipoprotein E-specific innate immune response in astrocytes from targeted replacement mice;** *J Neuroinflamm*, 3:10.
- Paszty C, Maeda N, Verstuyft J, Rubin EM. (1994) **Apolipoprotein AI Transgene Corrects Apolipoprotein E Deficiency-induced Atherosclerosis in Mice;** *J Clin Invest*, 94:899-903.
- Piedrahita, J.A., Zhang, S.H., Hagaman, J.R., Oliver, P.M., Maeda, N. (1992) **Generation of Mice Carrying a Mutant Apolipoprotein E Gene Inactivated by Gene Targeting in Embryonic Stem Cells,** *Proceedings of the National Academy of Science, USA*, Vol. 89, pp. 4471-4475.
- Plump A.S., Breslow, J.L. (1995) **Apolipoprotein E and the Apolipoprotein E-Deficient Mouse,** *Annual Review of Nutrition*, Vol. 15, pp. 495-518.
- Pratico, D., Tangirala, R.K., Rader, D.J., Rokach, J., FitzGerald, G.A. (1998) **Vitamin E Suppresses Isoprostane Generation in vivo and Reduces Atherosclerosis in ApoE-Deficient Mice,** *Nature Medicine*, Vol. 4, No. 10, pp. 1189-1192.
- Reddick, R.L., Zhang, S.H., Maeda, N. (1998) **Aortic Atherosclerotic Plaque Injury in Apolipoprotein E Deficient Mice,** *Atherosclerosis*, Vol. 140, No. 2, pp. 297-305.
- Reddick, R.L., Zhang, S.H., Maeda, N. (1994) **Atherosclerosis in Mice Lacking apoE. Evaluation of Lesional Development and**

Progression, *Atheroscl Thromb*, Vol. 14, No. 1, pp. 141-147.

Richardson, J.A., Burns, D.K. (2002) **Mouse Models of Alzheimer's Disease: A Quest for Plaques and Tangles**, *ILAR Journal*, Vol. 43, No. 2, pp. 89-99.

Shibata M, Yamada S, Kumar R, Calero M, Bading J, Frangione B, Holtzman DM, Miller CA, Strickland DK, Ghiso J, Zlokovic BV. (2000) **Clearance of Alzheimer's amyloid- β_{1-40} peptide from brain by LDL receptor-related protein-1 at the blood-brain barrier**. *Journal of Clinical Investigation*, 106(12):1489-99.

Sullivan, P.M., Mace, B.E., Maeda, N., Schmechel, D.E. (2004) **Marked Regional Differences of Brain Human Apolipoprotein E Expression in Targeted Replacement Mice**, *Neuroscience*, Vol. 124, pp. 725-733.

Tsukamoto, K., Tangirala, R., Chun, S.H., Pure, E., Rader, D.J. (1999) **Rapid Regression of Atherosclerosis Induced by Liver-Directed Gene Transfer of ApoE in ApoE-Deficient Mice**, *Arterioscler Thromb Vasc Biol*, Vol. 19, No. 9, pp. 2162-2170.

Tsukamoto, K., Heister, K.G., Smith, P., Usher, D.C., Glick, J.M., Rader, D.J. (1997) **Comparison of Human apoA-1 Expression in Mouse Models of Atherosclerosis After Gene Transfer Using a Second Generation Adenovirus**, *Journal of Lipid Research*, Vol. 38, pp. 1869-1876.

Yuan H, Wong LS, Bhattacharya M, Ma C, Zafarani M, Yao M, Schneider M, Pitas RE, Martins-Green M. (2007) **The effects of second-hand smoke on biological processes important in atherogenesis**. *BMC Cardiovascular Disorder*, 7:1.

Zhang, S.H., Reddick, R.L., Avdievich, E., Surles, L.K., Jones, R.G., Reynolds, J.B., Quarfordt, S.H., Maeda, N. (1997) **Paradoxical Enhancement of Atherosclerosis by ProbucoL Treatment in Apolipoprotein E-Deficient Mice**, *Journal of Clinical Investigation*, Vol. 99, No. 12, pp. 2858-2866.

Zhang S.H., Reddick, R.L., Burkey, B., Maeda N. (1994) **Diet-Induced Atherosclerosis in Mice Heterozygous and Homozygous for Apolipoprotein E Gene Disruption**, *Journal of Clinical Investigation*, Vol. 94, pp. 937-945.

Zhang S.H., Reddick, R.L., Piedrahita J.A., Maeda N. (1992) **Spontaneous Hypercholesterolemia and**

Arterial Lesions in Mice Lacking Apolipoprotein E, *Science*, Vol. 258, pp. 468-471.