

Transgenic HLA Models

MODELS DESIGNED TO TAKE YOUR STUDY FURTHER

Transgenic HLA Models

In vivo investigations of human immune system function can be frustrating when you don't have the right tools. Transgenic mice that express human HLA molecules represent a unique *in vivo* experimental model for evaluating human immune system function.

These models have been used to study the role of the human class I or class II restricted T cell repertoire in autoimmune disease, infectious disease, and vaccine development.

They are also valued tools for evaluating human HLA restricted T cell-mediated vaccine efficacy in oncology applications.

Taconic's collection of transgenic mice that express human HLA molecules have helped advance the science related to human immune system function, as well as potential novel therapeutics.

Check out Taconic's transgenic HLA models at Taconic.com/HLA



EXCLUSIVELY FROM TACONIC BIOSCIENCES

MODEL NUMBER	MODEL NAME
9662	HLA-A1 Mouse
9659	HLA-A2.1 Mouse
9660	HLA-A11 Mouse
9663	HLA-A24 Mouse
9661	HLA-B7 Mouse
9664	HLA-B44 Mouse
4149	Abb Knockout / Transgenic HLA-DR4 Mouse

KEY APPLICATIONS OF THE TRANSGENIC HLA MODELS

VACCINE RESEARCH

Transgenic HLA mice are great tools to help identify epitopes or to test vaccine efficacy in challenge studies, and support rational vaccine design through epitope identification (Ishioka et al. 1999, Okochi et al. 2008). The models can be used to study T cell responses restricted by HLA molecules (Alexander et al. 2003, Alexander et al. 1997, Okochi et al. 2008, Vitiello et al. 1991), and given the range of HLA types offered in these models, this facilitates the development of vaccines targeted against specific ethnic/regional populations, as well as broad populations. Models in this portfolio have been used in research on immunogenicity and vaccines directed against human lymphocytic choriomeningitis virus (LCMV) infection (Botten et al. 2007), HIV (McKinney et al. 2004, Nanjundappa et al. 2012, Singh et al. 2002, Wilson et al. 2003), hepatitis B virus (Depla et al. 2008, Oseroff et al. 1998) and Lassa virus (Botten et al. 2006), among others (Cong et al. 2011, Cong et al., 2012, Kotturi et al. 2009, Tan et al. 2010, Weiskopf et al. 2011).

ONCOLOGY

Transgenic HLA mice are used for cancer immunotherapy research. Example applications include evaluation of HLA restricted CTL epitope responses *in vivo*, determining the immunogenicity of novel cancer vaccines (Casnici et al. 2012, Hospers et al. 2009, Kang et al. 2013, Men et al. 1999, Tomita et al. 2013), and to test expansion and response of chimeric antigen receptor T cells.

Custom crosses to immunodeficient models or other genetically modified strains are available. Please contact your Strategic Account Manager for more information.

AUTOIMMUNE DISEASES

HLA models have proven to be important tools in the study of autoimmune disease, since certain HLA types are associated with autoimmune diseases. For example:

 HLA-DR4 is associated with rheumatoid arthritis and multiple sclerosis. The Abb Knockout/Transgenic HLA-DR4 Mouse expresses hybrid class II molecules with the peptide binding domains of human HLA-DRA and HLA-DRB*0401 and the membrane proximal domains of mouse I-E on an H2-Ab1 knockout background. These mice are susceptible to experimentally allergic encephalomyelitis (EAE) (Ito et al. 1996, Kawamura et al. 2008), as well as arthritis (Walker et al. 2012), and induced connective tissue disease (Greidinger et al. 2008).

INFECTIOUS DISEASES AND BIOTERROR AGENTS

Transgenic HLA models may be used to study human T cell response to infections to help elucidate if, and how, a protective response is initiated. For highly pathogenic emerging infections and bioterror agents, access to infected or immune human patients is sometimes not possible. Transgenic HLA models fill this important need. Use of predictive algorithms in combination with *in vivo* experiments in transgenic HLA mice permits epitope identification without having to manipulate dangerous pathogens, or having access to human donors. The models have been used in the study of arenaviruses (including Lassa, lymphocytic choriomeningitis, Guanarito, Junin, Machupo, Sabia and Whitewater Arroyo viruses) (Botten et al. 2010, Kotturi et al. 2009), HIV (McKinney et al. 2004, Nanjundappa et al. 2012, Singh et al. 2002, Wilson et al. 2003), poxviruses (Pasquetto et al. 2005), and hepatitis B (Oseroff et al. 1998).

IMMUNOGENICITY TESTING AND SAFETY ASSESSMENT

Transgenic HLA models might be exactly what you need when you are looking for a tool to screen immunogenicity of biologic drugs.

RELATED MODELS

Taconic offers other models which can be useful for these research applications.

Immunodeficient mice reconstituted with a human immune system via engraftment of human hematopoietic stem cells can be a complementary model to the transgenic HLA mice.

The CIEA NOG mouse® is the premier model for this type of reconstitution experiment and can be easily engrafted via a simple tail vein injection of CD34+ cord blood cells. Taconic now offers NOG mice which have been engrafted with human PBMCs or hematopoietic stem cells.

To learn more about the CIEA NOG mouse®, visit Taconic.com/NOG

TRANSGENIC HLA MOUSE MODELS

Alexander J, Oseroff C, Sidney J, Wentworth P, Keogh E, Hermanson G, Chisari FV, Kubo RT, Grey HM, Sette A. (1997) Derivation of HLA-A11/ Kb transgenic mice: functional CTL repertoire and recognition of human A11-restricted CTL epitopes. J Immunol 159(10):4753-61. Erratum in: J Immunol 1999 162(5):3104.

Alexander J, Oseroff C, Sidney J, Sette A. (2003) Derivation of HLA-B*0702 transgenic mice: functional CTL repertoire and recognition of human B*0702-restricted CTL epitopes. Hum Immunol 64(2):211-23.

Botten J, Alexander J, Pasquetto V, Sidney J, Barrowman P, Ting J, Peters B, Southwood S, Stewart B, Rodriguez-Carreno MP, Mothe B, Whitton JL, Sette A, Buchmeier MJ. (2006) Identification of protective Lassa virus epitopes that are restricted by HLA-A2. J Virol. 80(17):8351-61.

Botten J, Whitton JL, Barrowman P, Sidney J, Whitmire JK, Alexander J, Ting JP, Bui HH,

Sette A, Buchmeier MJ. (2007) HLA-A2restricted protection against lethal lymphocytic choriomeningitis. J Virol. 81(5):2307-17.

Botten J, Sidney J, Mothé BR, Peters B, Sette A, Kotturi MF. (2010) Coverage of related pathogenic species by multivalent and crossprotective vaccine design: arenaviruses as a model system.

Microbiol Mol Biol Rev. 74(2):157-70.

Botten J, Whitton JL, Barrowman P, Sidney J, Whitmire JK, Alexander J, Kotturi MF, Sette A, Buchmeier MJ. (2010) A Multivalent Vaccination Strategy for the Prevention of Old World Arenavirus Infection in Humans. J Virol. 84(19): 9947-56.

Cong H, Mui EJ, Witola WH, Sidney J, Alexander J, Sette A, Maewal A, McLeod R. (2011) Towards an immunosense vaccine to prevent toxoplasmosis: protective Toxoplasma gondii epitopes restricted by HLA-A*0201. Vaccine, 29(4):754-62. Cong H, Mui EJ, Witola WH, Sidney J, Alexander J, Sette A, Maewal A, McLeod R. (2012) Human immunome, bioinformatic analyses using HLA supermotifs and the parasite genome, binding assays, studies of human T cell responses, and immunization of HLA-A*1101 transgenic mice including novel adjuvants provide a foundation for HLA-A03 restricted CD8+ T cell epitope based, adjuvanted vaccine protective against Toxoplasma gondii. Immunome Res, 6:12.

Cong H, Mui EJ, Witola WH, Sidney J, Alexander J, Sette A, Maewal A, El Bissati K, Zhou Y, Suzuki Y, Lee D, Woods S, Sommerville C, Henriquez FL, Roberts CW, McLeod R. (2012) Toxoplasma gondii HLA-B*0702-restricted GRA7(20-28) peptide with adjuvants and a universal helper T cell epitope elicits CD8(+) T cells producing interferon-y and reduces parasite burden in HLA-B*0702 mice. Hum Immunol, 73(1):1-10.

Casnici C, Volpe G, Crotta K, Lattuada D, Saglio G, Marelli O. (2012) Immunologic evaluation of peptides derived from BCR/ABL-out-of-frame fusion protein in HLA A2.1 transgenic mice. J

Immunother. 35(4):321-8.

Depla E, Van der Aa A, Livingston BD, Crimi C, Allosery K, De Brabandere V, Krakover J, Murthy S, Huang M, Power S, Babé L, Dahlberg C, McKinney D, Sette A, Southwood S, Philip R, Newman MJ, Meheus L. (2008) Rational design of a multiepitope vaccine encoding T-lymphocyte epitopes for treatment of chronic hepatitis B virus infections. J Virol 82(1):435-50.

Dubey JP, Ferreira LR, Martins J, McLeod R. (2012) Oral oocyst-induced mouse model of toxoplasmosis: effect of infection with Toxoplasma gondii strains of different genotypes, dose, and mouse strains (transgenic, outbred, inbred) on pathogenesis and mortality. Parasitology, 139(1):1-13.

Fu Q, Wu Y, Yan F, Wang N, Wang W, Cao X, Wang Y, Wan T. (2011) Efficient induction of a Her2-specific anti-tumor response by dendritic cells pulsed with a Hsp70L1-Her2(341-456) fusion protein. Cell Mol Immunol. 8(5):424-32.

Greidinger EL, Zang YJ, Jaimes K, Martinez L, Nassiri M, Hoffman RW. (2008) CD4+ T cells target epitopes residing within the RNA-binding domain of the U1-70-kDa small nuclear ribonucleoprotein autoantigen and have restricted TCR diversity in an HLA-DR4transgenic murine model of mixed connective tissue disease. J Immunol. 180(12):8444-54.

Hospers GA, Meijer C, Dam WA, Roossink F, Mulder NH. (2009) Construction of a triple modified p53 containing DNA vaccine to enhance processing and presentation of the p53 antigen. Vaccine. 28(2):386-91.

Irwin MJ, Heath WR, Sherman LA. (1989) Species-restricted interactions between CD8 and the alpha 3 domain of class I influence the magnitude of the xenogeneic response. J Exp Med 170(4):1091-101.

Ishioka GY, Fikes J, Hermanson G, Livingston B, Crimi C, Qin M, del Guercio MF, Oseroff C, Dahlberg C, Alexander J, Chesnut RW, Sette A. (1999) Utilization of MHC class I transgenic mice for development of minigene DNA vaccines encoding multiple HLA-restricted CTL epitopes. J Immunol 1999, 162(7):3915-25.

Ito K, Bian HJ, Molina M, Han J, Magram J, Saar E, Belunis C, Bolin DR, Arceo R, Campbell R, Falcioni F, Vidovic D, Hammer J, Nagy ZA. (1996) HLADR4-IE chimeric class II transgenic, murine class II-deficient mice are susceptible to experimental allergic encephalomyelitis. J Exp Med 183(6):2635-44.

Kang YJ, Zeng W, Song W, Reinhold B, Choi J, Brusic V, Yamashita T, Munshi A, Li C, Minvielle S, Anderson KC, Munshi N, Reinherz EL, Sasada T. (2013) Identification of human leucocyte antigen (HLA)-A*0201-restricted cytotoxic T lymphocyte epitopes derived from HLA-DOβ as a novel target for multiple myeloma. Br J Haematol. 163(3):343-51.

Kawamura K, McLaughlin KA, Weissert R, Forsthuber TG. (2008) Myelin-reactive type B T cells and T cells specific for low-affinity MHC-binding myelin peptides escape tolerance in HLA-DR transgenic mice. J Immunol. 181(5):3202-11.

Kotturi MF, Botten J, Sidney J, Bui HH, Giancola L, Maybeno M, Babin J, Oseroff C, Pasquetto V, Greenbaum JA, Peters B, Ting J, Do D, Vang L, Alexander J, Grey H, Buchmeier MJ, Sette A. (2009) A multivalent and cross-protective vaccine strategy against arenaviruses associated with human disease. PLoS Pathog. 5(12):e1000695.

McKinney DM, Skvoretz R, Livingston BD, Wilson CC, Anders M, Chesnut RW, Sette A, Essex M, Novitsky V, Newman MJ. (2004) Recognition of variant HIV-1 epitopes from diverse viral subtypes by vaccine-induced CTL. J Immunol. 173(3):1941-50.

Men Y, Miconnet I, Valmori D, Rimoldi D, Cerottini JC, Romero P. (1999) Assessment of immunogenicity of human Melan-A peptide analogues in HLA-A*0201/ Kb transgenic mice. J Immunol 162(6):3566-73.

Nanjundappa RH, Wang R, Xie Y, Umeshappa CS, Xiang J. (2012) Novel CD8+ T cell-based vaccine stimulates Gp120-specific CTL responses leading to therapeutic and long-term immunity in transgenic HLA-A2 mice. Vaccine. 30(24):3519-25.

Okochi M, Hayashi H, Ito A, Kato R, Tamura Y, Sato N, Honda H. (2008) Identification of HLA-A24-restricted epitopes with high affinities to Hsp70 using peptide arrays. J Biosci Bioeng. 105(3):198-203.

Oseroff C, Sette A, Wentworth P, Celis E, Maewal A, Dahlberg C, Fikes J, Kubo RT, Chesnut RW, Grey HM, Alexander J. (1998) Pools of lipidated HTL-CTL constructs prime for multiple HBV and HCV CTL epitope responses. (2012) Vaccine, 16(8):823-33.

Pasquetto V, Bui HH, Giannino R, Banh C, Mirza F, Sidney J, Oseroff C, Tscharke DC, Irvine K, Bennink JR, Peters B, Southwood S, Cerundolo V, Grey H, Yewdell JW, Sette A. (2005) HLA-A*0201, HLA-A*1101, and HLA-B*0702 transgenic mice recognize numerous poxvirus determinants from a wide variety of viral gene products. J Immunol. 175(8):5504-15. Erratum in: J Immunol. 175(12):8440.

Singh RAK and Barry MA. (2002) Generation of Genome-Wide CD8 T Cell Responses in HLA-A*0201 Transgenic Mice by an HIV-1 Ubiquitin Expression Library Immunization Vaccine. J Immunol. 168(1):379-91.

Tan AC, La Gruta NL, Zeng W, Jackson DC. (2011) Precursor frequency and competition dictate the HLA-A2-restricted CD8+ T cell responses to influenza A infection and vaccination in HLA-A2.1 transgenic mice. J Immunol. 187(4):1895-902.

Tan TG, Mui E, Cong H, Witola WH, Montpetit A, Muench SP, Sidney J, Alexander J, Sette A, Grigg ME, Maewal A, McLeod R. (2010) Identification of T. gondii epitopes, adjuvants, and host genetic factors that influence protection of mice and humans. Vaccine, 28(23):3977-89.

Tomita Y, Yuno A, Tsukamoto H, Senju S, Kuroda Y, Hirayama M, Irie A, Kawahara K, Yatsuda J, Hamada A, Jono H, Yoshida K, Tsunoda T, Kohrogi H, Yoshitake Y, Nakamura Y, Shinohara M, Nishimura Y. (2013) Identification of promiscuous KIF20A long peptides bearing both CD4+ and CD8+ T-cell epitopes: KIF20Aspecific CD4+ T-cell immunity in patients with malignant tumor. Clin Cancer Res. 19(16):4508-20.

Vitiello A, Marchesini D, Furze J, Sherman LA, Chesnut RW. (1991) Analysis of the HLA-restricted influenza-specific cytotoxic T lymphocyte response in transgenic mice carrying a chimeric human- mouse class I major histocompatibility complex. J Exp Med 173(4):1007-15.

Walker KM, Rytelewski M, Mazzuca DM, Meilleur SA, Mannik LA, Yue D, Brintnell WC, Welch I, Cairns E, Haeryfar SM. (2012) Preventing and curing citrulline-induced autoimmune arthritis in a humanized mouse model using a Th2polarizing iNKT cell agonist. Immunol Cell Biol. 90(6):630-9.

Weiskopf D, Yauch LE, Angelo MA, John DV, Greenbaum JA, Sidney J, Kolla RV, de Silva AD, de Silva AM, Grey H, Peters B, Shresta S, Sette A. (2011) Insights into HLA-restricted T cell responses in a novel mouse model of dengue virus infection point toward new implications for vaccine design. J Immunol, 187(8):4268-79.

Wilson CC, McKinney D, Anders M, MaWhinney S, Forster J, Crimi C, Southwood S, Sette A, Chesnut R, Newman MJ, Livingston BD. (2003) Development of a DNA vaccine designed to induce cytotoxic T lymphocyte responses to multiple conserved epitopes in HIV-1. J Immunol 171(10):5611-23.

To view additional references for the Taconic Transgenic HLA Mouse Models, visit www.taconic.com/hla

© Taconic Biosciences, Inc. All rights reserved. Contents of this publication may not be reproduced in any form without prior permission.

US: 1-888-822-6642 EU: +45 70 23 04 05 INFO@TACONIC.COM

